

***Eugenia caryophyllata* Thunberg - A MIRACULOUS HERB**

ROMAN Luminița, ROMAN Horatiu, HOSU Anamaria, VASILIU Cristiana, MIHĂESCU Grigore, CZOBOR Ilida

Abstract. *Eugenia caryophyllata* Thunberg 1788 belongs to the family Myrtaceae, is an aromatic tree, originally from Indonesia used as a condiment. Its antibacterial, antifungal and anthelmintic properties are known from ancient times. Despite the development of the pharmaceutical industry, currently at least 30 000 people die every year in Europe due to infections caused by antibiotic-resistant microorganisms: *Staphylococcus aureus* resistant to methicillin (MRSA), *Staphylococcus aureus* vancomycin-resistant (VRSA), *Enterococcus spp.* vancomycin-resistant (VRE), *Streptococcus pneumoniae* penicillin-resistant (PRSP), Enterobacteriaceae (*Escherichia coli*, *Klebsiella pneumoniae*) resistant to the third generation cephalosporins, Enterobacteriaceae (*K. pneumoniae*) carbapenems-resistant and non-enteric bacteria (*Pseudomonas aeruginosa*) resistant to carbapenems. For almost every existing antibiotic, bacteria have developed a resistance factor that protects them from the action. For each resistance factor, pharmaceutical companies have developed a stronger antibiotic - until today. In the battle between bacteria and antibiotics the balance of victory begins to tilt toward these microorganisms. In these circumstances, the return to traditional medicine seems to be the solution. In order to determine the antibacterial activity of the compounds of *Eugenia caryophyllata* buds extract against bacteria isolated from nosocomial infections MDR (multiple drug resistant), we used the disc-diffusion method and inoculation of 96-well plate BHI. Screening of genes coding β -lactam antibiotics resistance was performed by PCR. The active compounds of the ethanol extracts and essential oil of *Eugenia caryophyllata* were determined by HPTLC, respectively by GC-MS. Gram positive and Gram negative strains isolated from nosocomial infections showed genotypic resistance characteristics to lactam antibiotics. Hydroalcoholic extracts and essential oil of *E. caryophyllata* were active against all MDR bacteria. Eugenol is the main component of the extract of *E. caryophyllata*. In conclusion, the extracts of *E. caryophyllata* can successfully replace antibiotics whose action against MDR bacteria proved to be ineffective.

Keywords: *Eugenia caryophyllata*, eugenol, MDR bacteria.

Rezumat. *Eugenia caryophyllata* Thunberg - o plantă miraculoasă. *Eugenia caryophyllata* Thunberg 1788, face parte din familia Myrtaceae, este un arbore aromatic, original din Indonezia folosit ca și condiment. Proprietățile sale antibacteriene, antifungice și antihelmitice sunt cunoscute din antichitate. În ciuda dezvoltării industriei farmaceutice, în prezent, cel puțin 30 000 de oameni mor anual în Europa datorită infecțiilor cauzate de microorganisme rezistente la antibiotice: *Staphylococcus aureus* rezistent la meticilină (MRSA), *Staphylococcus aureus* rezistent la vanomicină (VRSA), *Enterococcus spp.* rezistente la vanomicină (VRE), *Streptococcus pneumoniae* rezistent la penicilină (PRSP), enterobacterii (*Escherichia coli*, *Klebsiella pneumoniae*) rezistente la cefalosporine de generația a treia, enterobacterii (*K. pneumoniae*) rezistente la carbapeneme, și non-enterici (*Pseudomonas aeruginosa*) rezistente la carbapeneme. Pentru aproape fiecare antibiotic existent în prezent, bacteriile au dezvoltat un factor de rezistență care le protejează de acțiunea sa. Pentru fiecare factor de rezistență, companiile farmaceutice au dezvoltat un antibiotic mai puternic – până astăzi. În lupta dintre bacterii și antibiotice, balanța victoriei începe să se încline către aceste microorganisme. În aceste condiții, întoarcerea la medicina tradițională pare a fi soluția. Pentru a determina activitatea antibacteriană a compușilor din extract de muguri de *Eugenia caryophyllata* împotriva bacteriilor MDR izolate din infecții nosocomiale, am folosit metoda disc-difuzimetrică adaptată și însământare în plăci cu 96 godeuri cu mediu BHI. Screening-ul genelor codificatoare ale rezistenței la antibioticele β -lactamice s-a realizat prin metoda PCR. Compușii activi din extractele etanolice și ulei esențial din *Eugenia caryophyllata* au fost determinați prin HPTLC respectiv, GC. Tulipinile Gram pozitive și Gram negative izolate din infecții nosocomiale au prezentat caracteristicile genotipice de rezistență la antibioticele lactamice. Extractele hidroalcoolice și uleiul volatil din *E. caryophyllata* au avut activitate împotriva tuturor bacteriilor MDR. Eugenolul este componentul principal al extractului din *E. caryophyllata*. În concluzie, extractele din *E. caryophyllata* pot înlocui cu succes antibioticele a căror acțiune împotriva unor bacterii MDR s-a dovedit a fi ineficientă.

Cuvinte cheie: *Eugenia caryophyllata*, eugenol, bacterii MDR.

INTRODUCTION

The healing with medicinal plants is as old as mankind itself. The relationship between man and aromatic herbs dates back to the distant past, as it results from different sources: written documents, preserved monuments, and even the original drugs from plants (PETROVSKA, 2012). *Eugenia caryophyllata* Thunb, with EUCA15 symbol is a spice widely praised for its antibacterial and antioxidant properties, used for centuries as a food preservative. Currently, many articles in PubMed render the antibacterial, antifungal, antiviral and anticarcinogenic activity of the extracts of *E. caryophyllata* buds (clove). *E. caryophyllata* is an evergreen tree with a height of 10-20 m (TAJUDDIN et al., 2004). It is part of the Myrtaceae, being indigenous in India, Indonesia, Zanzibar, Mauritius and Sri Lanka. Buds of this tree are currently marketed worldwide. The production of flower buds begins 4 years after planting, being collected in the maturation phase before flowering (CORTÉS-ROJAS, 2014).

The classes of compounds with antimicrobial activity obtained from the extracts of *E. caryophyllata* are secondary metabolites derived from fundamental processes of photosynthesis, glycolysis and the Krebs cycle. Secondary metabolites in general are not essential for growth, development and reproduction of an organism and are either the result of the body's adaptation to external environmental factors or defense mechanisms against predator bodies, helping the survival of the organism. The changes of the biosynthetic pathways may be due to natural causes (for example, viruses or environmental changes), or unnatural causes (for example, chemical pollutants), in an effort to adapt to survive (DIAS, 2012). Plant extracts have multiple action target in the microbial cell, which is an advantage of their use as antimicrobials, with synergistic or

concerted activity of components - their antimicrobial activity, and not only as a result of the fact that they are multicomponent systems. This pharmacologic synergistic or additive effect may be beneficial by eliminating the side effects associated with the predominance of one of the xenobiotic compounds in the body.

This role of several chemicals to act synergistically or the additive myth originated in plant secondary compounds involved in the defense for the survival of the species. For example, role-products in the defense of a mixture of chemicals that have additive or synergistic effects on multiple target sites would not only ensure efficacy against a wide variety of herbivores or pathogens, but would also reduce the chances of these bodies to develop and adapt their reactions. Although secondary compounds may have a variety of functions in plants, it is likely that the functions can have an effect on the human body with medicinal potential. For example, cytotoxicity secondary compounds involved in plant defense against microbial pathogens could be useful as antimicrobial agents in humans (BRISKIN, 2000).

Ethanolic extracts and essential oil of clove have been shown to be an important source of phenolic compounds such as flavonoids, hydroxybenzoic acid, hydroxycinnamic acid and hydroxyphenyl propene. Eugenol is the main bioactive compound that was reported by different authors as having the largest activity (TAJUDDIN et al. 2004, JOSHI et al., 2010. BHOWMIK et al., 2010). Tannins were reported in large quantities: phenolic acids (caffeic acid, ferulic, ellagic and salicylic acid), gallic acid can be found in higher concentration. Flavonoids such as kaempferol, quercetin and its derivatives (glycosylated) have also lower concentrations. Another important compound in the essential oil of clove found in concentrations of up to 2.1% is α -humulen. Other volatile compounds present in low concentrations in the essential oil of cloves are β -pinene, limonene, farnesol, benzaldehyde, 2-heptanone, and ethyl hexanoate (CORTÉS-ROJAS, 2014).

MATERIAL AND METHODS

1. Preparation of the extract and analysis of the compounds

The buds of *Eugenia caryophyllata* were purchased in store. The ethanolic extract was prepared by macerating the clove powder in 95% ethyl alcohol for 24 hours (1: 4 w / v), after which it was introduced into a rotary evaporator for 10-15 minutes and then the supernatant was purified by Whatman filter paper no. 41. The extract thus obtained was stored in an amber glass container at 4°C. The analysis of the compounds of the ethanolic extracts was made using HPTLC (High Performance Thin-Layer Chromatography method). Chemicals and reagents use: 2,2 azinobis (3-ethylbenzothiazoline-6 sulfonic acid) diammonium salt (ABTS), ascorbic acid (vitamin C), gallic acid, Folin-Ciocalteu's reagent, and sodium persulfate ($\text{Na}_2\text{S}_2\text{O}_8$) were purchased from Merck (Darmstadt, Germany). Aluminium chloride (AlCl_3), sodium acetate (CH_3COONa), sodium carbonate (Na_2CO_3) and methanol were obtained from Chimopar (Bucharest, Romania). All chemicals and solvents were analytical grade. The absorbance measurements were performed using a T80+ UV/VIS Spectrophotometer (PG-Instruments). Each experiment has been performed in triplicate. For antioxidant activity of extracts we performed the following steps: the cationic radical ABTS⁺ was obtained from the reaction of 7 mmol/L ABTS diammonium salt solution with 2.45 mmol/L $\text{Na}_2\text{S}_2\text{O}_8$ solution mixed in 1:1 (v/v) ratio, incubated for 24 h at room temperature in the dark. Then, 0.5 mL of appropriately diluted extract were added to 3 mL ABTS⁺ solution diluted to achieve an absorbance less than 0.800, and the absorbance was measured at 734 nm, after 20 min. Calibration was performed using vitamin C as standard, in the concentration range of 50–250 $\mu\text{g}/\text{mL}$, following the same procedure.

The calibration curve was used to calculate antioxidant activity. For total phenolic content (TPC) determination, 1.5 mL of Folin-Ciocalteu reagent (0.2 mol/L) was added to 0.3 mL of plant extract appropriately diluted with distilled water. The reaction mixture was allowed to react 5 min and then, 1.2 mL of 0.7 mol/L Na_2CO_3 was added. The results were read using an absorbance spectrophotometry at 760 nm. A calibration curve was obtained using 0–100 mg gallic acid/mL and was used to calculate the total phenolic content of the extracts. The total flavonoids content of extracts was determined by treating 0.5 mL of extract appropriately diluted with distilled water with 0.4 mL of 25 g/L AlCl_3 solution, 0.5 mL of 100 g/L CH_3COONa solution and 4 mL distilled water. After 15 min, the absorbance of the mixture was measured at 430 nm and the flavonoid content express in mg rutin/g of plant, was calculated using the calibration curve obtained in the 0–120 mg/mL concentration range. The volatile oil of cloves was obtained by hydro distillation in a Clevenger-Neo for 4 h. The volatile oil was dried with Na_2SO_4 and stored in a dark glass bottle at 4°C. Oil samples were diluted in dichloromethane (1/200) for the analysis of the chemical compounds. To identify the compounds of the clove oil extract we used a GC-MS. The used gas chromatograph was Fisons Instruments GC 8000 coupled with mass spectrometer ionization quadruple analyzer impact, pattern MD 800. Ionization energy was 70 eV. Capillary column used was a fused silica column with 5% phenyl poly(dimethylsiloxane) (SLB-5ms, 30m x 0.32mm id, film thickness of stationary phase 0.25 μm). The operating conditions were: split splitless injector (injection mode - split flow dividing ratio 1/30) to 280°C, ion source temperature 200°C and 280°C interface; initial column temperature was 40°C scheduled as follows: 3 min at 40°C, 4°C / min up to 280°C, isothermal conditions (at 280°C) for 20 min; the flow rate of the carrier gas (helium) was 2 ml / min; injected sample volume was 1 mL. Data acquisition was performed with the software MassLab in Table 30-600 amu, with a scan rate of 1 scan / s. Identification of the detected compounds was based on comparing their mass spectra with those in databases (NIST, WILEY). Relative concentration of the compounds was calculated using the values chromatographic peak areas under the curves, without applying correction factors.

2. Analysis of resistance factors MDR strains from nosocomial infections

Strains isolated from urogenital infections were from patients hospitalized at Theodor Burghel Hospital Bucharest. Phenotypic resistance to antibiotics was determined by disk diffusion method. For the analysis of virulence factors we used

phenotypic tests that allow the detection of six extracellular virulence factors: hemolysis, caseinase, gelatinase and pore-forming toxins: DN-ase, lipase and lecithinase. These extracellular proteins known as the invasins have a role in the degradation of tissue and invasion. Highlighting these soluble factors of virulence was achieved by growing strains on culture media and studying the specific detection of each type of virulence factor. β -lactamases are the most important enzymes that confer antibacterial resistance. To identify genes β -lactam antibiotic resistance we used PCR method. Preparation of bacterial DNA to screen for gene was made by the method of bacterial lysis. The strains were seeded on an agar plate and kept at 37 °C for 24 hours. In the PCR tubes there were added 20 μ l NaOH (0.05 M) and SDS (0.25%), which made suspensions of 1-5 colonies. The tubes bacterial suspensions were kept for 15 minutes at 95°C, after which there were added 180 ml extraction of TE solution (10 mM Tris-HCl, pH 8, 0.1 mM EDTA), followed by centrifugation at 13,000 rpm / min for 3 minutes. The supernatant was recovered, and then stored in a refrigerator. The verification of the product obtained was carried out by electrophoresis of 0.8% agarose gel stained with ethidium bromide 3.5 μ g / ml. Following observation of bacterial phenotypes it was checked the existence of several genes commonly associated with the production of β -lactamases: *bla_{CTX-M}*, *bla_{NDM}*, *bla_{OXA}*, *bla_{IMP}*. For the amplification of this gene, we used the primer sequences described in the literature.

Table 1. Sequences of the primers used in the PCR reaction.

Gene	Primers sequence (5' to 3', as synthesized)	Expected amplicon size (bp)	Reference
<i>bla_{OXA-48}</i>	OXA-48 F: GCG TGG TTA AGG ATG AAC AC OXA-48 R: CAT CAA GIT CAA CCC AAC CG	438 bp	Poirel et.al., 2004
<i>bla_{TEM}</i>	TEM F: ATA AAA TTC TTG AAG ACG AAA TEM R: GTC AGT TAC CAA TGC TTA ATC	1080 bp	Eftekhari et.al., 2005
<i>bla_{CTX-M}</i>	CTX-M F: CGC TGT TGT TAG GAA GTG TG CTX-M R: GGC TGG GTG AAG TAA GTG AC	730 bp	Schlesinger et.al., 2005
<i>bla_{NDM}</i>	NDM F: GGT TTG GCG ATC TGG TTT TC NDM R: CGG AAT GGC TCA TCA CGA TC	621 bp	Nordmann et.al., 2011
<i>bla_{VIM}</i>	VIM F: CAG ATT GCC GAT GGT GTT TGG VIM R: AGG TGG GCC ATT CAG CCA GA	523 bp	Docquier et.al., 2003
<i>bla_{IMP}</i>	IMP F: GGA ATA GAG TGG CCT AAT TCT IMP R: GTG ATG CGT CYC CAA YTT CAC	361 bp	Docquier et.al., 2003

3. Determination of antimicrobial activity of extracts of clove

The quantitative determination of antimicrobial activity and establishing the minimum inhibitory concentration (MIC) was made by the method of serial microdilution in a liquid medium BHI (Hearth Infusion Broth) in 96well plates / Ependorf tubes of 1.5ml (CHIFIRIUC et. Al., 2011). MIC was established macroscopically, as the last concentration at which no growth of the microbial environment was observed and that the appearance of turbidity are read spectrophotometrically to the absorbance at 620 nm. For the qualitative testing of the antimicrobial activity, there were prepared microbial suspension adjusted to 1.5x10⁸ CFU / mL of the 0.5 McFarland standard, 15-18 hour cultures grown on solid medium (Sabouraud). Antimicrobial activity was determined by disc diffusion method adapted to standardized control antimicrobial activity of antibiotics, CLSI, 2009. The microbial suspension, adjusted to 0.5 McFarland standard in the cloth was seeded in the cloth agar Petri dishes which pipetted 10 μ L of the stock solution obtained of the plant extract. The plates are incubated for 16-18 hours at 35 ± 2° C, with the lid down. Reading the results was done by measuring the diameters of the zones of inhibition comparatively. The influence on the ability adhesion to the inert substrate was quantified after the protocol quantitative analysis of the effect of antimicrobial, evaluating the biomass, after fixation with methanol and staining with crystal violet.

The optical density was determined spectrophotometrically and the biological material resuspended at 490 nm (NORUZI et. al., 2010). For determining the inhibitory activity of clove extracts against soluble virulence factors, the strains were grown in increasing concentrations of stock solutions subinhibitory essential oil: DMSO (CMI / 2) and the positive control (untreated strain) were used for seeding each medium with each specific culture for detecting soluble virulence factor (hemolysins caseinase, lipase, lecithinase and DN-ase).

RESULTS AND DISCUSSION

Table 2. From the analysis of the ethanol extract of cloves, HPTLC method has shown the following results:

Ethanol extract	Antioxidant activity (mg vitamin C / g of herbs)	Total polyphenol content (mg gallic acid / g herbs)	The content of flavonoids (mg rutin / g herbs)
<i>Eugenia caryophyllata</i>	4.16087	264.44	0.33

It is a very remarkable antioxidant, so a gram of ethanol extract clove corresponds to 4160.87 g of vitamin C. It is also important to note the high concentration of polyphenols (264.44 mg gallic acid / g herbs). However, the concentration of flavonoids is almost insignificant. The concentration of polyphenolic compounds in herbs is coupled with their antioxidant activity. Antioxidant properties are induced by redox activity and mechanisms of action such as: free-radical scavenging activity, transition-metal-chelating activity and/or singlet-oxygen-quenching capacity. Polyphenols have an important role in the stabilization of lipid peroxidation and inhibit various types of oxidative

enzymes. The relationship between total antioxidant activity and the phenolic content of a large number of spices has not been investigated sufficiently. Many researchers have argued that the phenolic compounds in spices were responsible for their antioxidant activity, but few could correlate these relations on the basis of statistical data (SHAN et al., 2005; DE ROSA & CRUTCHLEY, 2002).

The oil obtained by hydrodistillation of cloves was intense yellow with characteristic aromatic and persistent odour. The average yield clove oil extraction was 8.40%, the density was 1.055 (g/cm³) and the index of fraction was 1.535. In clove oil there were identified five compounds representing approximately 95.51% of the total. In diagram 1 are shown the major component is eugenol, present in an amount of 87%, followed by caryophyllene (4.87%), acetyl eugenol (2.11%), β -pinene (1.021%) and hexadecanoic acid, octadecyl ester (0.51%).

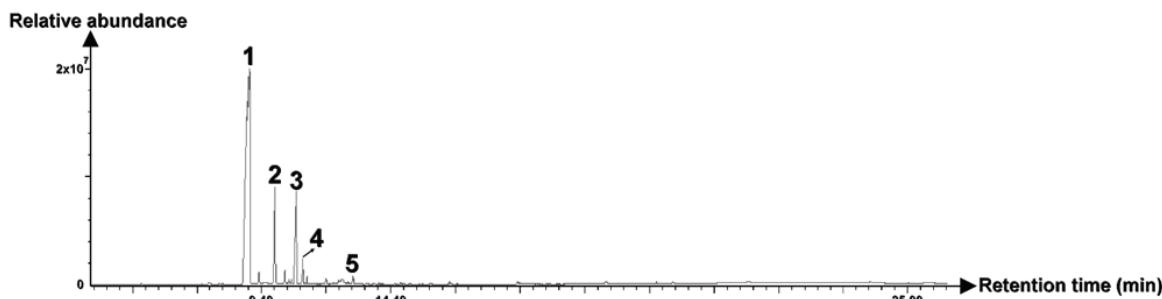


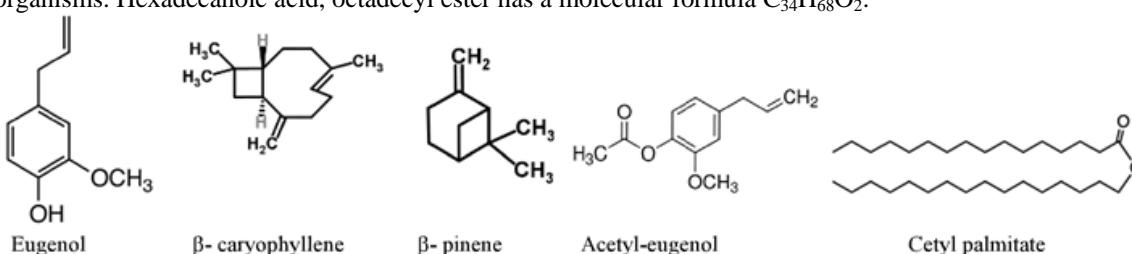
Diagram 1. Chromatogram of the essential oil of clove: the abscissa is the last retention time (minutes) and the ordinate, the relative abundance. Compounds are listed in order of elution: 1=eugenol, 2=caryophyllene, 3=acetyl eugenol, 4= β -pinene, 5=hexadecanoic acid, octadecyl ester.

Eugenol or phenol, 2-methoxy-3-(2-propenyl) has the molecular formula C₁₀H₁₂O₃ and a molecular weight of 164 201 Da. Eugenol is an aromatic compound, the ether-oxide group with methylenedioxy group attached to the benzene nucleus in para-position to the allyl chain. Eugenol increased the permeability of the membrane; the deformation of macromolecules in the membrane was verified by spectroscopy (DEVI et al., 2011). The compound is a very promising candidate for versatile applications; eugenol possesses significant antioxidant and has also been used as a penetration enhancer (PRAMOD et al. 2010). Eugenol is also known for his property against microorganisms forming biofilms. Adherence of the microorganisms to host cells and tissues is the first event required for initial colonization or establishment of infection. Moreover, the microbial surface contact can trigger various cellular behaviours, including biofilm formation. Biofilms can be defined as irreversibly surface-attached communities of cells (sessile cells) embedded in a self-produced exopolymeric matrix, displaying a distinctive phenotype compared to their free-floating (planktonic cells) counterparts. Sessile cells are less susceptible to medicaments. Biofilms are thereby difficult to eradicate (DE PAULA et al., 2014, CHIFIRIUC et al., 2011). Eugenol is polar due to the acidic hidroxyl (OH) group, but acetyl-eugenol is not polar. As a result, they can be separated by extraction from a 5% aqueous NaOH solution. Acetyl-eugenol will dissolve in the organic CH₂Cl₂ layer, while eugenol remains in the aqueous base layer as a phenoxide.

Caryophyllene is a bicyclic sesquiterpene, usually found as a mixture of isocaryophyllene (cis double bond isomer) and α - humulen (obsolete name: α -caryophyllene), an open-ring isomer. Caryophyllene is considered to be a rare structure in nature, a cyclobutane ring (GERTSCH et al., 2008) with molecular formula C₁₅H₂₄. CALLEJA et al., 2014, reported the antimicrobial activity of caryophyllene. β -pinene, a monoterpene, is one of the two isomers of pinene; the other one is α -pinene. It is soluble in alcohol but not in water. It smells of green wood. It is also known as 6,6-dimethyl-2-methylenebicyclo [3.1.1] heptane. Bactericidal and bacteriostatic activity of β -pinene has been reported in many articles. β -pinene has a direct effect on the respiratory system of fungi by inhibiting the efflux pump protons, K⁺ and H⁺ that are found in the mitochondrial membrane, since the loss of respiratory control and the disintegration of organelles followed by an inhibition of breathing at higher concentrations of terpenes . β - pinene has a preferential affinity for mitochondria (URIBE et al., 1985).

Acetyl-eugenol or phenol, 2-methoxy-4-(2-propenyl) - acetate has a molecular formula C₁₂H₁₄O₃.

Hexadecanoic acid, octadecyl ester or cetyl palmitate is the ester derived from palmitic acid and cetyl alcohol. Palmitic acid, in IUPAC nomenclature, is the most common fatty acid (saturated) found in animals, plants and microorganisms. Hexadecanoic acid, octadecyl ester has a molecular formula C₃₄H₆₈O₂.



Following the comments of phenotypic resistance to antibiotics by disc-diffusion method, 70% of strains were resistant to extended spectrum β -lactams (ceftazidime, cefotaxime, aztreonam), 54% were resistant to clavulanic acid, 41% were resistant to carbapenems (meropenem, imipenem) and 39% were resistant to oxacillin.

The results of the analysis of soluble factors revealed the presence of exotoxins and pore-forming toxins. 7 strains showed simultaneous four soluble virulence factors, 9 strains presented three virulence factors simultaneously and 12 showed two virulence factors simultaneously. Of the 82 studied strains, following phenotypic tests for evidence of virulence factors, 56 strains showed proteases by testing simple agar medium with the addition of milk. The test was considered positive when observed around colonies yellowish-white precipitate of calcium paracaseinase. In this paper 20 strains of all studied strains showed gelatinase on average egg yolk medium with added lecithin seed purified. In the case of positive reaction around the bacterial colony emerged crop or a halo and / or opacified area. Clarification result from the release of more soluble lecithin of lipid complex (presence of lecithinase). 18 strains showed lipase on agar medium with the addition of TWEEN 80 (sorbitol monooleate). The presence of TWEEN-lipase crystals oleate led to the emergence Ca^{2+} insoluble (crystal formed between fatty acids and the release of Ca^{2+} ions). Of the total studied strains, 17 strains showed hemolysins on agar medium with addition of 5% sheep blood or rabbit blood (highlighting the presence of Kanagawa hemolysins). Viewing the occurrence of areas of haemolysis around the colonies, as a transparent halo, the clear characteristic was β -haemolytic strains (which achieve a complete haemolysis; the haemoglobin released from red blood cells by the action of α -hemolysins generates a green halo, emphasizing partial degradation). Of the total strains, 16 strains showed DN-aze on agar medium with DNA with toluidine blue. DNA hydrolysis was revealed by the appearance of a halo around the spot pink culture, environment while the rest remained blue coloured. Of the 82 studied strains, 21 showed gelatinase nutrient agar medium with gelatine. After precipitation around the colonies a halo appeared due to the hydrolysis of gelatine.

For screening the activity of oil clove against soluble virulence factors, the strains were seeded in BHI broth medium with the addition of the clove oil extract diluted in DMSO (1: 2) as the minimum inhibitory. Thus, the obtained strains were passaged in specific medium of virulence factors. The inhibitory effect of the extracts of clove against virulence factors has been reported in a ratio of 100%. According to the assessment, the phenotypical resistance factor β -lactam antibiotics, there have been selected primers for molecular screening of the major coding for the β -lactamase gene described in the literature as being frequent (PATERSON & BONOMO, 2005, LIVERMORE, 1995). For β -extended spectrum lactamases in the case of bacteria resistant to penicillins and cephalosporins there have been used *bla*_{CTX-M} and *bla*_{TEM} genes encoding for β -lactamases CTX-M. and TEM. For metal- β -lactamase we used *bla*_{VIM}, *bla*_{KPC} and *bla*_{OXA-23} genes coding for VIM, KPC and OXA-23 carbapenemases (BUSH & JACOBY, 2010). Figure 1 shows the negative image of ethidium bromide-stained agarose gel of the multiplex PCR assay of *bla*_{TEM-like} and *bla*_{CTX-M-like} for strains investigated.

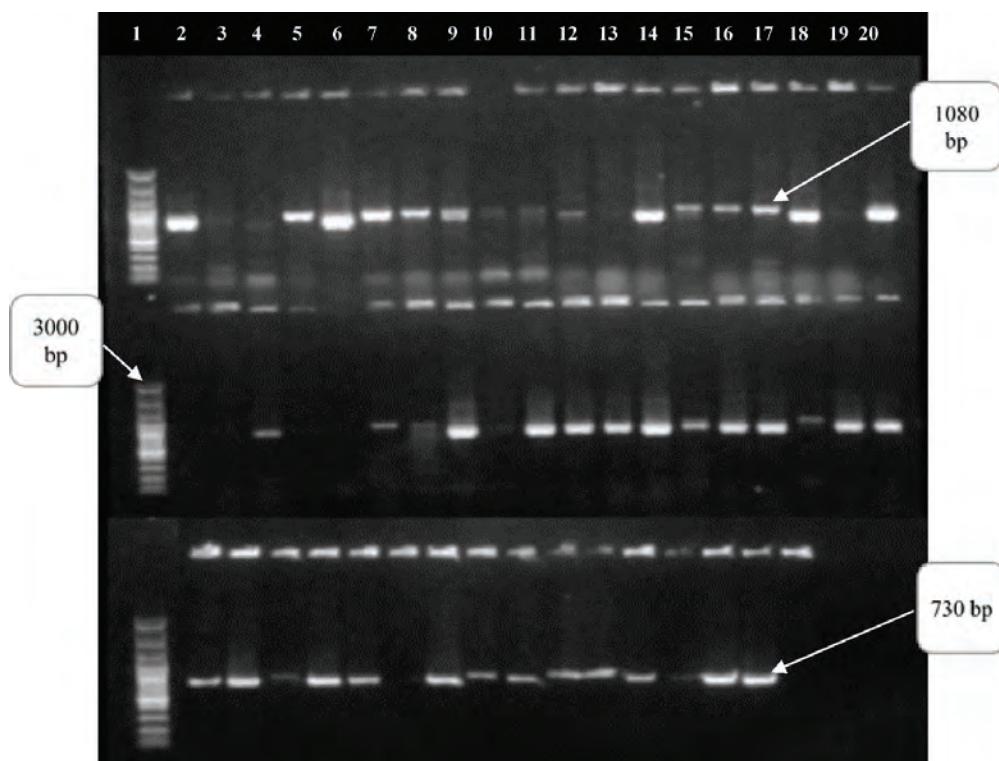


Figure 1 Agarose gel electrophoresis by multiplex PCR amplification for simultaneous detection of *bla*_{TEM-like} and *bla*_{CTX-M-like} genes, numbered lanes correspond to the following samples: top row(1-20): molecular weight markers Gene Ruler 3000bp (Fermentas) (MGM), *E.coli*₇, *E.coli*₂₉, *E.coli*_{2R}, *E.coli*₁₃, *E.coli*_{3R}, *E.coli*₂₂, *E.coli*₁₉, *E.faecalis*₄, *E.faecalis*_{7R}, *E.faecalis*₂, *E.coli*₂₈, *E.coli*₁₀₂₂, *E.coli*₃₆, *E.coli*₁₂, *E.coli*₁₀₁₈, *E.coli*₁₀₁₀₁, *E.coli*₁₁, *E.coli*₃₃, *E.coli*_{17R}; middle row (1-20): (MGM), *E.coli*_{24I}, *E.coli*₂₆, *E.coli*₅, *E.coli*₁₁₃₈, *E.coli*_{13R}, *E.coli*₃₂, *E.coli*₁₀₂, *E.coli*_{0R}, *A.baumannii*_{22R}, *E.coli*_{16R}, *K.pneumoniae*_{6R}, *K.pneumoniae*_{3R}, *E.coli*₁₁₂₉₃₇, *K.pneumoniae*_{2R}, *K.pneumoniae*_{15R}, *K.pneumoniae*₂₀, *E.faecalis*₄₁, *K.pneumoniae*_{19I}, *K.pneumoniae*_{12R}; bottom row: *K.pneumoniae*_{9III}, *E.coli*₈₁, *K.pneumoniae*_{26I}, *Proteus miserabilis*_{1I}, *K.pneumoniae*_{14I}, *K.pneumoniae*_{3II}, *E.coli*₁₁₃₈, *K.pneumoniae*₁₁₃₇₄₀, *K.pneumoniae*_{10I}, *E.coli*_{5III}, *K.pneumoniae*_{K1}, *K.pneumoniae*_{K2}, *K.pneumoniae*_{K3}, *E.coli*_{E1}, *E.coli*_{E2}, *E.coli*_{E3}; (B): simultaneous detection of *bla*_{NDM-like} and *bla*_{OXA-48-like} genes: (MGM), *K.pneumoniae*_{6I}, *K.pneumoniae*_{14I}, *K.pneumoniae*_{15I}, *K.pneumoniae*_{19I}, *K.pneumoniae*₀.

Amplicon approx. 1080 bp corresponding bla_{TEM} gene confirming the presence of β -lactamases TEM can be observed in the wells 5, 7, 8, 9, 15, 16, 17 (in the top row), in the wells 7, 8, 18 (in the middle row) and the wells 4, 9, 11, 12 in the lower row (representing 26% of the 54 strains). Amplicons approx. 754 bp corresponding bla_{CTX-M} gene confirming the presence of β -lactamases CTX-M can be observed in the wells 2, 6, 14, 18, 20 (in the top row), in the wells 4, 9, 11, 12, 13, 14, 15, 16, 17, 19, 20 (in the middle row) and in the wells 2, 3, 5, 6, 8, 10, 13, 14, 15, 16 (in bottom row). Molecular analysis showed that the 26% of strains showed the presence of β -lactamases TEM-type compared with 48% of the strains showed β -lactamase CTX-M type. The multiplex PCR method did not reveal the presence of bla_{VIM}, bla_{IMP}, bla_{NDM} and bla_{OXA-48} genes coding for VIM, IMP, NDM and OXA-48 carbapenemases. The other amplicons are not detected although not ruling out nonspecific β -lactamase. The existence of these types of β -lactamases confirms lactam antibiotics resistance to pathogens.

Qualitative and quantitative evaluation of the antimicrobial activity of clove extracts

The qualitative testing by disc diffusion method Kirby-Bauer, adapter (CHIFIRIUC et al., 2011) emphasized that hydroalcoholic extracts and essential oil of *E. caryophyllata* shoots had antimicrobial activity for all bacterial strains from the collection achieved, quantified by the appearance of a zone of growth inhibition around the spot stock solution deposited on the seeded agar medium as can be seen in the example from Figure 2.

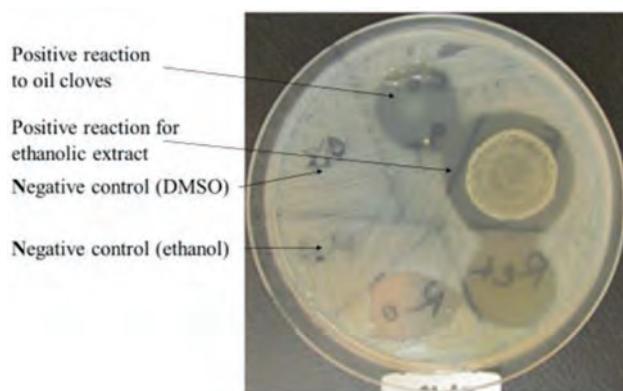


Figure 2. Qualitative testing of susceptibility to volatile oil and ethanol extract of *E. caryophyllata* strains studied, for example *Pseudomonas aeruginosa* III. (original).

Qualitative evaluation is an estimate, because it is not known the extent of absorption of the compound by the medium. Quantitative determination of antimicrobial activity was performed by the technique of successive binary microdilution in a liquid medium, which is a method of determining the minimum inhibitory concentration (MIC). The assay was performed according to the method described by CHIFIRIUC et al., 2011. Out of stock solutions of the compounds to be analysed there are performed micro binary successive dilutions in liquid BHI medium, distributed in 96-well plates. No dilutions were made in the wells treated for positive control (culture medium) and negative control (the control microbial culture).

MIC values are between 7.8 and 62.5 μ L/mL for ethanolic extracts and essential oils of *E. caryophyllata*. In Table 2, there are shown the MIC values for Gram negative strains from the collection of study. Considering some of the compounds resulting from the secondary metabolism of plants, also have a protective role, for example against phytopathogens, which are generally Gram-negative bacteria, we can make a correlation between these phytopathogenic bacteria and pathogenic bacteria in humans. Thus, we could explain the action of active compounds from plants against a broad spectrum of pathogens.

Table 2. The minimal inhibitory concentrations (MICs) for the essential oil and ethanolic extracts of *E. caryophyllata* against Gram negative bacteria.

Strain	MIC ₅₀ ethanolic extract (μ L/ml)		MIC ₅₀ essential oil (μ L/ml)	
	MIC ₅₀	MIC ₉₀	MIC ₅₀	MIC ₉₀
<i>Escherichia coli</i>	15,625	62,5	7,8	31,25
<i>Klebsiella pneumoniae</i>	62,5	125	7,8	31,25
<i>Proteus mirabilis</i>	31,25	62,5	31,25	7,8
<i>Alcaligenes faecalis</i>	31,25	62,5	15,625	31,25
<i>Acinetobacter baumannii</i>	15,625	62,5	7,8	15,625
<i>Pseudomonas aeruginosa</i>	31,25	62,5	7,8	31,25

CONCLUSIONS

Ethanol extracts and the essential oil of clove can be used successfully for severe nosocomial infections, due in particular to MDR Gram negative bacteria. Eugenol (the main component of *E. caryophyllata*) and the concerted action of the compounds resulting from secondary metabolism of *E. caryophyllata* had through several targets inhibitory action against Gram negative bacteria. The importance of isolating synthetic compounds in the extracts of cloves can be materialized by obtaining of new drugs for treatment of infections caused by MDR bacteria. Promoting the use of plant extracts is the main aim to eradicate microorganisms whose virulence is due to the abuse of drugs. The use of spices in daily consumption, especially cloves, for a long-term therapy, will result in increased body immunity against pathogens.

ACKNOWLEDGEMENS

We express our gratitude to Professor Coordinator, Mihăescu Grigore, Mrs. Carmen Maria Chifiriuc director of the Research Institute of the Faculty of Biology, Mrs. Dr. Camelia Tudor, head of the laboratory of Theodor Burghel Hospital Bucharest.

REFERENCES

- BHOWMIK D., KUMAR K. P. S., YADAV A., SRIVASTAVA SHWETA, PASWAN S., DUTA AMIT SANKAR. 2012. Recent Trends in Indian Traditional Herbs *Syzygium aromaticum* and its Health Benefits. *Journal of Pharmacognosy and Phytochemistry*. **1**(1): 13-23 <http://www.phytojournal.com/vol1Issue1/1.html> (Accessed: December 26, 2014).
- BRISKIN D. P. 2000. Medicinal plants and phytomedicines. Linking plant biochemistry and physiology to human health. *Plant Physiology*. **124**: 507-514. <http://www.plantphysiology.org/content/124/2/507.short> (Accessed: March 21, 2015).
- BUSH K. & JACOBY G. A. 2010. Updated functional classification of β -lactamases. *Antimicrobial Agents and Chemotherapy*. **54**(3): 969-976. <http://www.ncbi.nlm.nih.gov/pubmed/19995920>. (Accessed: March 12, 2015).
- CALLEJA M. A., VIEITES J. M., MONTERO-MELÉNDEZ T., TORRES M. I., FAUS M. J., GIL A., SUÁREZ A. 2014. The antioxidant effect of β -caryophyllene protects rat liver from carbon tetrachloride-induced fibrosis by inhibiting hepatic stellate cell activation. *British Journal of Nutrition*. **109**(3): 394-401. <http://www.ncbi.nlm.nih.gov/pubmed/22717234> (Accessed: March 21, 2015).
- CHIFIRIUC MARIA CARMEN, MIHĂESCU G., LAZĂR VERONICA. 2011. Microbiologie și Virologie Medicală. Edit. Universității din București. 189-230.
- CORTÉS-ROJAS D. F., FERNANDEZ DE SOUZA CLAUDIA REGINA, OLIVEIRA W. P. 2014. Clove (*Syzygium aromaticum*): a precious spice. *Asian Pacific Journal of Tropical Biomedicine*. **4**(2): 90-96. <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3819475> (Accessed: March 08, 2015).
- DAS A. D., URBAN SYLVIA, ROESSNER U. 2012. A historical overview of natural products in drug discovery. *Metabolites*. **2**: 303-336. <http://www.mdpi.com/2218-1989/2/2/303/htm> (Accessed: February 26, 2015)
- DE PAULA SUELEN BALERO, BARTELI THAIS FERNANDA, DI RAMIO VANESSA, SANTOS PERERIA JUSSECANIA, MOREY T. A., BOSINI MARIANA ANDREA, NAKAMURA C. V., YAMAUCHI LM. AMADA-OGATTA SUEL FUMIE. 2014. Effect of eugenol on cell surface hydrophobicity, adhesion, and biofilm of *Candida tropicalis* and *Candida dubliniensis* isolated from oral cavity of HIV-infected patients. *Evidence-Based Complementary and Alternative Medicine*. 2014. 8 pp. <http://www.hindawi.com/journals/ecam/2014/505204/> (Accessed: March 15, 2015).
- DE ROSA MARIA C. & CRUTCHLEY R. J. 2002. Photosensitized singlet oxygen and its applications. *Coordination Chemistry Reviews*. (Accessed March 21, 2015). **233** (234): 351-371. <http://fisicaspecialistica.unipr.it/didattica/att/e279.4100.file.pdf> (Accessed March 21, 2015).
- DEVI K. P., NISHA S. A., PANDIAN S. K. 2011. Eugenol (an essential oil of clove) acts as an antibacterial agent against *Salmonella typhi* by disrupting the cellular membrane. *Journal of Ethnopharmacol*. **130**(1): 107-15. <http://www.researchgate.net/publication/43534149> (Accessed March 21, 2015).
- DOCQUIER J. D., RICCIO M. L., MUQNAIOIL C., LUZZARO F., TONILOLO A., AMISOSANTE G., ROSSOLINI G. M. 2003. IMP-12, a new plasmid-encoded metallo- β -lactamase from a *Pseudomonas putida* clinical isolate. *Antimicrobial Agents Chemother*. **47**(5): 1522-1528. <http://www.ncbi.nlm.nih.gov/pubmed/12709317> (Accessed: March 11, 2015).
- EFTEKHAR F., HOSSEINI-MAZINANI S. M., GHANDILI S., HAMRAZ M., ZAMANI S. 2005. PCR detection of plasmid mediated TEM, SHV and AmpC β -lactamases in community and nosocomial urinary isolates of *Escherichia coli* *Iranian Journal of Biotechnology*. **3**(1): 48-54. http://ijbiotech.neoscriber.com/?page=article&article_id=12951 (Accessed Jan 03, 2015)
- GERTSCH J., LEONTI M., RADUNER S., RAKZ I., CHEN J. Z., XIE X. Q., ALTMAN K. H., KARSAK A., ZIMMER A. 2008. Beta-caryophyllene is a dietary cannabinoid. *Proceedings of the National Academy of Sciences of the United States of America*. **105**(26): 9099-9104.

- <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC2449371> (Accesed: March 10, 2015).
- JOSHI B., SAH G. P., BASNET B. B., BHATT M. R., SHARMA D., SUBEDI K., PANDEY J., MLLA R. 2010. Phytochemical extraction and antimicrobial properties of different medicinal plants: *Ocimum sanctum* (Tulsi), *Eugenia caryophyllata* (Clove), *Achyranthes bidentata* (Datiwan) and *Azadirachta indica* (Neem). *Journal of Microbiology and Antimicrobials.* **3**(1): 1-7. 9
- http://www.academicjournals.org/article/article1380016710_Joshi%20et%20al.pdf (Accesed: March 11, 2015).
- LIVERMORE D. M. 1995. β -Lactamases in laboratory and clinical resistance. *Clin. Microbiology Review.* **8**(4):557-584. <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC172876/pdf/080557.pdf> (Accesed: March 11, 2015).
- NORMAN PATRICE, POIREL L., CARRÉR AMELIÉ, TOLEMAN M. M. A., WALSH T. R. 2011. How to detect NDM-1 producers. *Journal of Clinical Microbiology.* **49**(2): 718-721.
- <http://www.ncbi.nlm.nih.gov/pubmed/21123531> (Accesed: March 10, 2015).
- NOROUZI FATEMEH, MANSOURI SHAHLA, MORADI M., RAZAVI M. 2010. Comparison of cell surface hydrophobicity and biofilm formation among ESBL-and non-ESBL-producing *Pseudomonas aeruginosa* clinical isolates. *African Journal of Microbiology Research.* **4**(11): 1143-1147.
- <http://www.academia.edu/5066629> (Accesed: February 18, 2015)
- PATERSON D. L. & BONOMO R. A. 2005. Extended-spectrum β -lactamases: a clinical update. *Journal of Microbiology.* **18**: 657-686. <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC1265908> (Accesed: March 21, 2015).
- PETROVSKA B. B. 2012. Historical review of medicinal plants' usage. *Pharmacognosy Review.* **6**(11): 1-5. <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3358962> (Accesed: February 15, 2015).
- POIREL P., HÉRITIER C., TOLÜN V., NORMAND P. 2004. Emergence of oxacillinase-mediated resistance to imipenem in *Klebsiella pneumoniae*. *Antimicrobial Agents and Chemotherapy.* **48**(1):15–22. <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC310167/> (Accesed Jan. 03, 2015).
- PRAMOD K., ANSARI S. H., ALI J. 2010. Eugenol: a natural compound with versatile pharmacological actions. *Natural Product Communications.* **5**(12): 1999-2006. <http://www.ncbi.nlm.nih.gov/pubmed/21299140> (Accesed: March 10, 2015).
- SCHLESINGER J., NAVON-VENEZIA S., CHMELNITSKY I., HAMMER-MÜNZ O., LEAVIT A., GOLD H. S., SCHWABER M. J., CARMELI Y. 2005. Extended-spectrum beta-lactamases among *Enterobacter* isolates obtained in Tel Aviv, Israel. *Antimicrobial Agents and Chemotherapy.* **49**(3): 1150–1156. <http://www.biomedcentral.com/content/pdf/1476-0711-11-19.pdf> (Accesed Aug. 04, 2015).
- SHAN B., CAI Y. Z., SUN M., CORKE H. 2005. Antioxidant Capacity of 26 Spice extracts and characterization of their phenolic constituents. *Journal of Agricultural and Food Chemistry.* **53**(20): 7749-7759. <http://www.ncbi.nlm.nih.gov/pubmed/16190627> (Accesed: March 02, 2015).
- TAJDDUIN AHMAD S., LATIF A., QASMI I. A. 2004. Effect of 50% ethanolic extract of *Syzygium aromaticum* (L.) Merr. & Perry. (clove) on sexual behaviour of normal male rats. *BMC. Complementary and Alternative Medicine.* **4**: 17 pp. <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC534794> (Accesed: March 02, 2015).
- URIBE S., RAMIREZ J., PENÁ A. 1985. Effects of β -pinene on yeast membrane functions. *Journal of Bacteriology.* 1195-1200 pp. <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC215026>. (Accesed: March 02, 2015).
- ***. <https://en.wikipedia.org/wiki/Caryophyllene>. (Accesed: March 02, 2015).

Roman Luminița¹, Roman Horațiu², Hosu Anamaria³, Vasiliu Cristiana⁴, Mihăescu Grigore¹, Czobor Ilda¹

¹Faculty of Biology, University of Bucharest,

²Faculty of Geology, University of Bucharest,

³Faculty of Chemistry and Chemical Engineering Babes Bolyai Cluj,

⁴Social worker, London

E-mail: luminitaroman9@yahoo.com, horace_the_horace@yahoo.com,
hosuanamaria@yahoo.com, grigoremihăescu2006@yahoo.com

Received: March 27, 2015

Accepted: May 12, 2015